

C/N 5183

1100054334

Perpustakaan Sultanah Nur Zahirah (UMT)
Universiti Malaysia Terengganu

LP 6 FMSM 2 2007



1100054334

In-vitro genotoxic effects of copper in tilapia fingerlings (*Oreochromis niloticus*) / Harmeeta Kaur.

PERPUSTAKAAN SULTANAH NUR ZAHIRAH
UNIVERSITI MALAYSIA TERENGGANU (UMT)
21030 KUALA TERENGGANU

100054334

Iihat caholoh



**IN-VITRO GENOTOXIC EFFECT OF COPPER IN NILE TILAPIA
FINGERLINGS (*Oreochromis niloticus*)**

By

Harmeeta Kaur d/o Terlok Singh

**Research Report submitted in partial fulfillment of
The requirements for the degree of
Bachelor of Science (Marine Science)**

**Department of Marine Science
Faculty of Maritime Studies and Marine Science
UNIVERSITI MALAYSIA TERENGGANU**

2007

1100054334

This project report should be cited as:

Harmeeta, K. 2007. In-Vitro Genotoxic Effect of Copper in Tilapia fingerlings (*Oreochromis niloticus*). Undergraduate Thesis, Bachelor of Science (Marine Science), Faculty of Maritime Studies and Marine Science, Universiti Malaysia Terengganu.

No part of this project report may be reproduced by any mechanical, photographic or electronic process, or in the form of photographic recording, nor may it be stored in a retrieval system, transmitted or otherwise copied for public or private use, without written permission from the author and the supervisor of the project.



**JABATAN SAINS MARIN
FAKULTI PENGAJIAN MARITIM DAN SAINS MARIN
UNIVERSITI MALAYSIA TERENGGANU**

**PENGAKUAN DAN PENGESAHAN LAPORAN
PROJEK PENYELIDIKAN I DAN II**

Adalah ini diakui dan disahkan bahawa laporan penyelidikan bertajuk:

In-Vitro Genotoxic Effect of Copper in Tilapia Fingerlings (*Oreochromis niloticus*) oleh **Harmeeta Kaur a/p Terlok Singh**, No. matrik: **UK10092** telah diperiksa dan semua pembetulan yang disarankan telah dilakukan. Laporan ini dikemukakan kepada Jabatan Sains Marin sebagai memenuhi sebahagian daripada keperluan memperolehi **Ijazah Sarjana Muda Sains (Sains Samudera)** Fakulti Pengurusan Maritim dan Sains Marin, Universiti Malaysia Terengganu.

Disahkan oleh:

Penyelia Utama

Nama:

Cop Rasmi:

Tarikh: *May 6, 2007*

PROF MADYA DR. MOHD. EFFENDY ABD WAHID
Pengarah

Institut Bioteknologi Marin
Universiti Malaysia Terengganu
21030 Kuala Terengganu, Terengganu.

Ketua Jabatan Sains Marin

Nama: **DR. RAZAK ZAKARIYA**
Ketua Jabatan Sains Marin
Cop Rasmi: Fakulti Pengajian Maritim dan Sains Marin
Universiti Malaysia Terengganu
(UMT)

Tarikh: *9/1/08*

ACKNOWLEDGEMENT

Supreme thanks to God for permitting me to successfully complete this thesis. Warmest gratitude to my supervisor, Assoc. Prof. Dr. Effendy Abdul Wahid for his contribution and precious suggestions. Also, my special thanks to Dr. Nor Antonina Abdullah for her kind support and encouragement that I receive from her during my research.

Deepest sincere gratitude goes to Mr. Vijayendran Govindasamy for his guidance, advices and full time monitoring and had provided me valuable information throughout the completion of this research. I would also like take this opportunity to thanks Mr. Prem Kumar for helping me from the beginning till the end of this research. Thanks to science officers in INOS for their supports.

Not forgetting my parents Mr. Terlok Singh and Mrs. Balbir Kaur for their boundless support and encouragement which kept me motivated. In a nutshell, thanks to everyone who were involved directly or indirectly in the achievement of this research.

LIST OF CONTENTS

ACKNOWLEDGEMENT	ii
LIST OF CONTENTS	iii
LIST OF TABLES	vi
LIST OF FIGURES	vii
LIST OF APPENDICES	ix
LIST OF ABBREVIATIONS	xi
ABSTRACT	xii
ABSTRAK	xiv
CHAPTER 1 INTRODUCTION	1
CHAPTER 2 LITERATURE REVIEW	5
2.1 Nile Tilapia Fish (<i>Oreochromis niloticus</i>)	5
2.2 Median Lethal Concentrations (96-h LC ₅₀) Acute Toxicity Test	7
2.3 Gills, Muscle-tissue and Viscera of Fish	8
2.4 Heavy Metal	9
2.4.1 Copper	9
2.5 Comet Assay	11
2.6 DNA	12
2.7 Randomly Amplified Polymerase Chain reaction (RAPD-PCR)	13
2.7.1 Primers	15
CHAPTER 3 MATERIALS AND METHODS	
3.1 Background of Study Area	16
3.2 Toxicity Test Experimental Design	16

3.2.1	Experimental Materials	16
3.2.2	Water Source	17
3.2.3	Fish Source	17
3.2.4	Copper Stock Solution	18
3.2.5	Experimental Design of Acute Toxicity Test	18
3.2.6	Statistical Analysis	19
3.2.7	Experimental Design for Sub-lethal Toxicity Test	19
3.3	ICP-MS Analysis	22
3.3.1	Sample Preparation	22
3.3.2	Open Acid Digestion	22
3.3.3	Detection of Heavy Metal through ICP-MS	23
3.3.4	Calculation and Statistical Analysis	23
3.3.5	Recovery Test	23
3.4	Genome Study	24
3.4.1	Materials	24
3.4.2	DNA Isolation	24
3.4.3	Purification and Quantification of DNA	25
3.4.4	Primer	26
3.4.5	DNA Amplification by RAPD-PCR	26
3.4.6	Database Establishment	27
3.4.7	Data Analysis	27
3.5	Single Cell Gel Electrophoresis Study (Comet Assay)	28
3.5.1	Materials	28
3.5.2	Protocols	28
3.5.3	Microscope Examination	29
3.5.4	Statistical Analysis	30

CHAPTER 4 RESULTS

4.1	Part A: Determination of 96-hLC ₅₀ value and detection of copper accumulation concentrations in exposed fingerlings through ICP-MS	31
4.1.1	Physico-chemical Parameters	31
4.1.2	96-hLC ₅₀ Value of Copper	32
4.1.3	Intensity Graph of ICP-MS Analysis	33
4.1.4	Metal Recovery Test	34
4.1.5	Mean Copper Accumulation in Exposed Nile Tilapia Fingerlings for 7, 14 and 21 days	34
4.2	Part B: Genome Analysis of Exposed Nile Tilapia Fingerlings by Using RAPD-PCR	41
4.2.1	DNA Purification and Quantification	41
4.2.2	Random Amplified Polymorphic DNA (RAPD) – PCR	44
4.2.3	Cluster Analysis	49
4.2.4	Genomic DNA Template Stability	53
4.3	Part C: Investigating the Degree of Nucleus Damage in Cell through Single Gel Electrophoresis	57
CHAPTER 5 DISCUSSION		60
CHAPTER 6 CONCLUSION		77
REFERENCES		79
APPENDICES		83
CURRICULUM VITAE		104

LIST OF TABLES

Table		Page
4.1	Measured physico-chemical parameters of each test water	33
4.2	The calculated 96-h LC ₅₀ value	33
4.3	Concentrations design for continuous flow- through test	34
4.4	Metal recovery percentage for Cu ²⁺	35
4.5	Mean Cu accumulation in gills, stomach, tissue and whole fish of exposed tilapia fingerlings	36
4.6	Concentration and quantity of extracted DNA at 260 nm/ 280 nm absorbance ratio	42
4.7	Distributions in DNA tail percentage values	58

LIST OF FIGURES

Figure		Page
2.1	Nile tilapia fish (<i>Oreochromis niloticus</i>)	6
3.1	Overview diagram of continuous flow through system	21
3.2	Design of continuous flow-through system	21
4.1	Calibration Graph for ICP-MS Analysis	33
4.2	Cu uptake in gills of exposed Nile tilapia fingerlings	36
4.3	Cu uptake in viscera of exposed Nile tilapia fingerlings	37
4.4	Cu uptake in whole body of exposed Nile tilapia fingerlings	38
4.5	Cu uptake in muscle-tissue of exposed tilapia fingerlings	39
4.6	Overview of Cu accumulation in gills, viscera, muscle-tissue and whole body in of exposed Nile tilapia fingerlings for 7, 14 and 21 days	40
4.7	The electrophoresis pattern of tilapia fingerlings genomic DNA for 7 days exposure	42
4.8	The electrophoresis pattern of tilapia fingerlings genomic DNA for 14 days exposure	43
4.9	The electrophoresis pattern of tilapia fingerlings genomic DNA for 21 days exposure	43
5.0	RAPD profiles amplified by primer OPA 9 of Nile tilapia fingerlings	44
5.1	RAPD profiles amplified by primer OPB 7 of Nile tilapia fingerlings	45
5.2	RAPD profiles amplified by primer OPB 5 of Nile tilapia fingerlings	46
5.3	RAPD profiles amplified by primer OPB 8 of Nile tilapia fingerlings	47

5.4a	Dendogram cluster analysis with Nei & Li's Coefficient index based on RAPD patterns using primer OPA 9	49
5.4b	Dendogram cluster analysis with Nei & Li's Coefficient index based on RAPD patterns using primer OPB 5	50
5.4c	Dendogram cluster analysis with Nei & Li's Coefficient index based on RAPD patterns using primer OPB 7	51
5.4d	Dendogram cluster analysis with Nei & Li's Coefficient index based on RAPD patterns using primer OPB 8	52
5.5a	Genomic DNA template stability via RAPD-PCR by primer OPB 7	53
5.5b	Genomic DNA template stability via RAPD-PCR by primer OPB 5	53
5.5c	Genomic DNA template stability via RAPD-PCR by primer OPA 9	54
5.5d	Genomic DNA template stability via RAPD-PCR by primer OPB 8	54
5.6	Effects of different Cu concentrations on Nile tilapia fingerlings and DNA tail percentage for 7, 14 and 21 days	57
5.7	Representative images of normal and damaged cells after running the comet assay	59

LIST OF APPENDIXES

Appendix		Page
I	Dry and wet weight of stomach	83
II	Dry and wet weight of whole tilapia body	84
III	Dry and wet weight of gills	85
IV	Dry and wet weight of tissues	86
V	Body and total body length of Nile tilapia fingerlings	87
VI	Similarity coefficients on DNA fingerprints using primer OPB 7	88
VII	Similarity coefficients on DNA fingerprints using primer OPA 9	89
VIII	Similarity coefficients on DNA fingerprints using primer OPB 5	90
IX	Similarity coefficients on DNA fingerprints using primer OPB 8	91
X	RAPD fingerprinting patterns of tilapia using primer OPB 7	92
XI	RAPD fingerprinting patterns of tilapia using primer OPA 9	92
XII	RAPD fingerprinting patterns of tilapia using primer OPB 5	93
XIII	RAPD fingerprinting patterns of tilapia using primer OPB 8	93
XIV	ANOVA analysis for Cu ²⁺ concentrations in gills with Tukey's comparisons	94

XV	ANOVA analysis for Cu ²⁺ concentrations in whole body with Tukey's comparisons	96
XVI	ANOVA analysis for Cu ²⁺ concentrations in stomach with Tukey's comparisons	98
XVII	ANOVA analysis for Cu ²⁺ concentrations in tissues with Tukey's comparisons	100
XVIII	ANOVA analysis for Cu ²⁺ concentrations in cell (Single cell gel electrophoresis) with Tukey's comparisons	102

LIST OF ABBREVIATIONS

Cu	-	Copper
Cu^{2+}	-	Copper in ionic form
ppb	-	parts per billion
ppm	-	parts per million
HNO_3	-	Nitric acid
HCL	-	Hydrochloric acid
H_2O_2	-	Hydrogen Peroxide
$^{\circ}\text{C}$	-	degree Celsius
g	-	gram
mg	-	milligram
cm	-	centimeter
μg	-	microgram
mg L^{-1}	-	miligram per liter
μl	-	microliter
%	-	percentage
bp	-	base pair

ABSTRACT

The effects of prolonged exposure of copper through sub-lethal toxicity test using Nile tilapia (*Oreochromis niloticus*) fingerlings was investigated using five nominal copper concentrations (control, 0.1819, 0.3638, 0.7276 and 1.0914 mg L⁻¹). Throughout 21 days of exposure, tilapia fingerlings were harvested at 7, 14 and 21 days for mean copper accumulation detection through ICP-MS, investigation of cell damage through Single Cell Gel Electrophoresis and determining DNA structure alteration through RAPD-PCR analysis. The 96-h LC₅₀ value for Nile tilapia fingerlings was calculated to be 1.819 mg L⁻¹. Accumulation of copper was studied on four different parts; muscle-tissue, viscera, gills and whole fingerling body. The uptake of copper varied in an order from highest concentrations; viscera > whole body > gills > muscle-tissue. Highest Cu²⁺ concentration accumulated by viscera was 690.4904 µg g⁻¹ at 21 days. At cell level, damage induced by Cu²⁺ in tilapia fingerlings showed a significant increase in DNA tail percentage in 7 and 14 days of exposure, indicating DNA damage was observed at all concentrations compared with control ($P<0.05$). A gradual decrease in the mean DNA tail percentage was observed at 21 days indicating repair of the damage DNA. The mean DNA tail percentage showed a dose-related increase and time dependent decrease after the treatment with Cu²⁺ when compared to control. RAPD pattern displayed some changes in polymorphism band patterns in exposed Nile tilapia fingerlings. The disappearance of bands was found in highest Cu²⁺ concentration (1.0914 mg L⁻¹) for primer OPB 7 at band molecular size of 500 bp in 14 and 21 days of exposure. Disappearance of bands proved that DNA fragment of exposed tilapia were altered signifying that DNA damage had

occurred. Therefore, results in the present study proved that Cu²⁺ is significantly known as genotoxic compound to Nile tilapia fingerlings as damage in cell level and DNA fragment alteration was detected.

ABSTRAK

Kesan Genotoksik Kuprum Ke Atas Anak-Anak Ikan Nile Tilapia (*Oreochromis niloticus*)

Kesan pendedahan logam kuprum melalui ujian ketoksikan kronik yang menggunakan anak-anak ikan Nile tilapia (*Oreochromis niloticus*) telah dikaji dengan menggunakan lima jenis kepekatan logam kuprum (kawalan, 0.1819, 0.3638, 0.7276 and 1.0914 mg L⁻¹). Sepanjang pendedahan selama 21 hari, anak-anak ikan tilapia dibunuh pada hari ketujuh, keempat belas and hari kedua puluh satu untuk menentukan jumlah logam kuprum yang telah terakumulasi melalui analisis ICP-MS, menyiasat kerosakan yang berlaku pada peringkat sel melalui kaedah Single Cell Gel Electrophoresis dan menentukan perubahan yang berlaku pada struktur DNA melalui analisis RAPD-PCR. Nilai 96-h LC₅₀ bagi ujian penentuan ketoksikan akut logam kuprum telah dikira sebagai 1.819 mg L⁻¹. Akumulasi logam kuprum telah diselidik menggunakan empat bahagian berlainan iaitu tisu, visera, insang dan seluruh badan anak ikan tilapia. Jumlah akumulasi logam kuprum yang dianalisis, didapati mengikut turutan tersebut; visera > seluruh badan > insang > tisu. Kepekatan logam kuprum yang paling tinggi dikesan pada visera iaitu sebanyak 690.4904 µg g⁻¹ pada hari kedua puluh satu. Kerosakan yang disebabkan oleh Cu²⁺ pada bahagian sel ikan tilapia telah menunjukkan peningkatan dalam peratus ekor DNA pada hari ketujuh dan keempat belas, menandakan kerosakan DNA wujud untuk semua kepekatan Cu²⁺ berbanding dengan sampel kawalan ($p<0.05$). Penurunan secara beransur-ansur dalam peratus ekor DNA juga telah diperhatikan pada

hari kedua puluh satu, menandakan pemulihan berlaku pada DNA yang rosak selepas dianalisis melalui Single Cell Gel Electrophoresis. Kehilangan jalur dikesan pada kepekatan logam Cu^{2+} (1.0914 mg L^{-1}) bagi primer OPB 7 untuk jalur molekul yang bersaiz 500 bp di hari keempat belas dan kedua puluh satu. Kehilangan jalur membuktikan bahawa struktur DNA telah mengalami perubahan. Oleh itu, keputusan dari kajian ini membuktikan bahawa Cu^{2+} sudah terbukti sebagai agen genotoksik terhadap anak-anak ikan tilapia kerana kerosakkan pada peringkat sel dan pada struktur DNA dapat dikesan.